Analysis Of Routine Bacterial Contamination Checking Feedback Of Nasiriyah City Hospitals During 2016

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Abstract

Hospital contamination is an important cause of nosocomial infection. This study aims to assess the occurrence of bacterial contamination in 70 different departments of two private and five public hospitals in Nasseriah city (south of Iraq) during 2016. From 50 to 200 swabs collected weekly from each wards, were isolated 213 positive culture bacterial isolates, comprised 11 bacterial species detected by biochemical and enzymatic tests. The contaminant species (30%) to *Micrococcus* spp. and *Citrobacters* pp were the fewest (0.46%). Generally, the private hospitals were less contaminated than public ones, the winter and summer months were less contaminated than spring and autumn, and multi-specialty hospital have more contamination than single-specialty one.

Introduction

Nosocomial infections (also called Healthcare-Associated Infections HAIs) are defined as infections that occurred during a hospitalization and are not present prior to hospital attendance (1). Therefore, the hospital contamination have been recognized as a crucial problem affecting the quality of provided health care that can leads to increasing of patients' morbidity and mortality, length of hospital staying and elevate costs of health care.

Despite some normal flora do not cause a threat to healthy hospital staff, but may cause serious problem to some patients, other virulent species (such as *Klebsiella* spp, E. coli, *Enterobacter* spp., *Citrobacter* spp., *Acinetobacter* spp), that frequently shed by patients, contaminate surfaces for days, whereupon they increase acquisition risk for all other hospitalized patients (2-4).

One critical aspect of surfaces contamination spreading, is the ability of the pathogenic bacteria to survive for prolonged enough time to make it difficult to eradicate by cleaning and disinfection (5), increases therisk of infection transmission to a susceptible patient or healthcare worker (6).Therefore, previous many studiesdocumented the persistence of various bacterial species in thehospital inanimate environment for broad range period of time, for example: Acinetobacter spp. may be detected within 3 days to 5 months, E. coli from 1.5 h to over 30 months, Enterococcus spp. from 5 days to 4 months, and for S. aureus from 7 days to 7 months (7) and which may be carried in symptom-less the nasal passages of 30- 60% of Web Site: <u>https://imed.utq.edu.iq</u>

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personnel carriers. Pseudomonas survived for a months in wet environment, but only from a few hours to a few days on dry surfaces(3). Also, the bacterial features that influence the resistance against disinfectant and ultimately enhance survival rate, varies among bacterial species.Whereas Grambacteria negative have an outer membrane acting as a barrier preventing the uptake of disinfectants, some able to survive by biofilms formation or by virtue of their higher concentration on surfaces(8).

The contamination through direct contact occurs via cross-transmission and dissemination with contaminated inanimate surfaces (9). For example: Pseudomonas and spp Stenotrophomonas spp can adhere to biofilm of surfaces that protecting them from chlorine-containing and other types of disinfectants (10). Rogues et al. reported that 14% of ICU health care workers hands were Pseudomonas positive when washed with contaminated tap water and 12% were positive when the last contact was with a Pseudomonas positive patient (11), make it have a greater propensity to cause contamination.

E. coli and *Enterobacter* spp. are fecal bacteria and this suggests that stool of patients and hospital staff, urinary tract infected patients, food and water supply may be sources of fecal contamination. infection Acinetobacter spp was described in twooutbreaks in (12, 13), which infected more than 30 patients and 60 patients in ICUs and in CCU respectively. While, suchstudiesrevealed the role of frequently handled clinical equipment andmandatory glove use for outbreak reservoir staff were an (14). Other studies attributed the causes

of the spread of bacterial contamination to common hospital materialsparticularly those taken from patient to patient such as cotton, propylene plastic, and polyester(3) • immobileobjects like: stainless steels. plastic and computerkeyboards (15, 16)differences offurniture materials in hospital wards affected the adhesion of bacteria thatmakethe survival rate depending on the material made. Thus, the evaluation of surface bacterial contamination is useful in checking the effectiveness of thedisinfecting and cleaning practicesto prevent microbial colonization on hospital surfaces.

To achieve such challenge, the objective of our study is to analysesthe routine potentially pathogenic bacterial data contamination of selected departments of various responsibility and bed capacity hospitals, with special emphasis characteristics of isolated bacteria and whether there are differences among hospitals in relation to their responsibility, capacity and annual seasons.

Material and methods

The study was conducted from January 2016 to December 2016. Aweekly sample collection was done by taking of 50-200 with cotton-tipped swabs. The surfaces ofroutinely used equipmentand furniture of seventy wardsin Nasseriah hospitals, were examined. The placesthat suggested beingpotentially more prone to patients acquiring infection of hospital such as surgical operation halls, CCUs, ICUs.emergency rooms.birth hall,Preterm infantsrooms, blood exchange room, ENT units, ophthalmic unit were included. Routine culturing

was done directly by inoculationeach swab on three culturing media: nutrient agar, MacConkey agar, and blood agar by streaking method. The inoculated were incubated aerobically plates overnight at 37 °C for 24 hours, no anaerobic cultures were performed.Primary isolated strainswere examinedby Gram stain. colony morphology on culture media, and

tentative diagnosis by several biochemical reactions like enzymatic activity, carbohydrates fermentation tests, haemolysis, pigment production, mucous secretion and swarming phenomena. 70 departments of seven hospitals (2 private and 5 public) are involved in this study .Excel office 2006 program was used for statistical analysis of data and

generate illustrated figures.

Results

Investigation of bacterial contamination in hospitals during the whole year resulted in 213 bacterial positive cultures, classified within 11 species. In April we obtained the highestcount of isolated bacterial strains from the largest number of hospital facilities and achieved the highest percentage of contamination, while in July the previous results were the lowest (as shown in Figure 1). There was tendency towards decreased bacterial contamination inboth the summer and winter seasons in comparison with increased of incidence of positive culture in spring and autumn seasons, (as shown in Figure 2).

Identification of bacterial species revealedthat *Pseudomonas*spp was the most frequentlyisolated bacteria(64 time, 29.9%), followed by Enterobacterspp(41 time, 19.15%), *E. coli* (36 time, 16.82%), *Staph aureus*(26 times, 12.14%), Bacillus spp(17 times, 7.94%), *Acinetobacter*(14 times, 6.54%), *Stenotrophomonas*spp(9 times, 4.2%),*Klebsiellas*pp(3 times, 1.4%), *Burkholderiacepacia*(2 times, 0.93%),while *Citrobacter*spp and *Micrococcus* spp were the fewer contaminant bacterial strains (1 time, 0.46%), (as shown in figure 3 and 4).

In terms of annual bacterial strain count, private hospitals were less contamination (only one bacterial strain for each) than public hospitals (Figure 5). To achieve a fair level of comparison, we divided the number of studied department in each hospital by the total number of bacterial isolates obtained from to measure the level of contamination (see figure 6). Overall, the results showed improper levels of required disinfecting and cleaning, especially in the most crowded and reviewed governmental hospitals in the region.

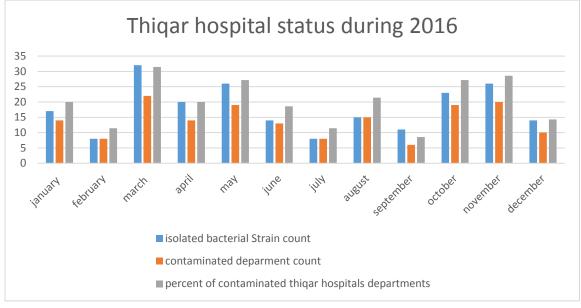


Figure 1: Nasseriah hospital state during 2016: illustrated as isolated bacterial strain count (blue column), number of contaminated departments (orange column) and percent of contaminated departments (grey column) as a part of 70 studied one.

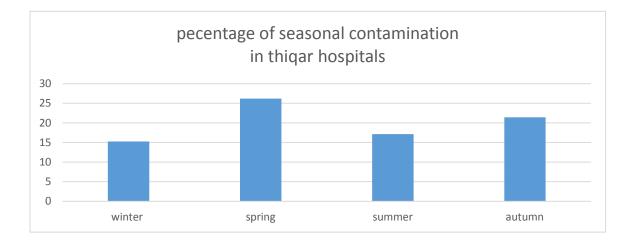


Figure 2: Seasonal emergence of contaminant bacteria in Nasseriah hospitals: duringspring and autumn, Nasseriah Hospitals have more contamination than other seasons.

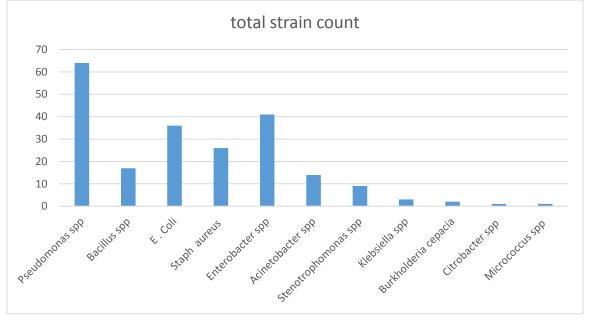
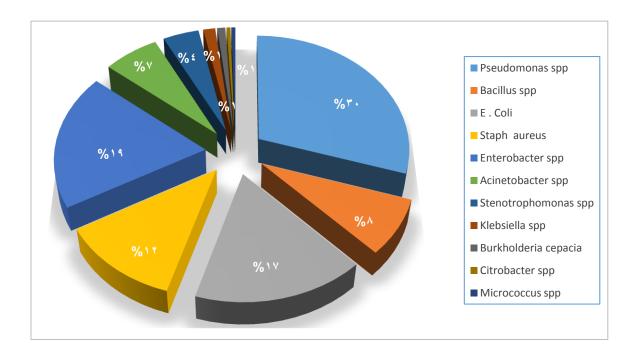


Figure 3: Bacterial species prevalence in Nasseriah hospitals: illustrated as frequency of bacterial isolation



Thi-Qar Medical Journal (TQMJ):Vol.(17),No.(1),2019 Web Site: https://imed.utq.edu.iq ISSN (Print):1992-92 18, ISSN (Online):1992-92 18 DOI: https://doi.org/10.32792/utq/utjmed/17/1/14/0 Figure 4: Percentage of isolated bacterial species from tot

Figure 4: Percentage of isolated bacterial species from total 214 isolates. Pseudomonasspp was the more prevalence strain while, Citrobacter and Micrococcus were more rare contaminant ones.

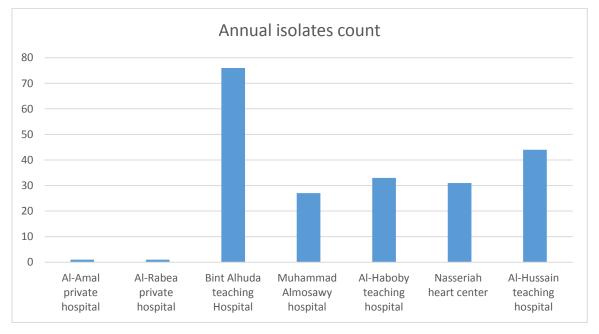


Figure 5: Annual isolated bacterial strain count of Nasseriah hospitals: total bacterial strains isolated from different departments of city hospitals. Clearly the public hospitals were more bacterial contaminated than private one.

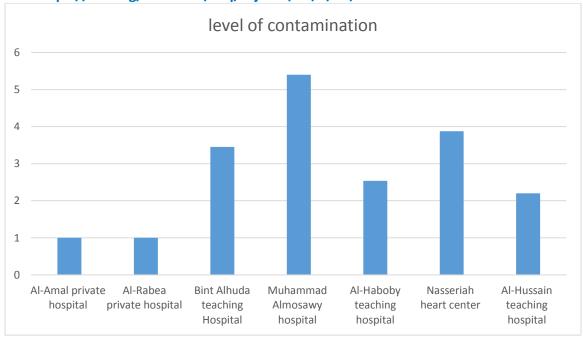


Figure 6: Level of contamination of Nasseriah hospitals: the level is calculated by division of number of examined departments of each hospital by the number of isolated bacterial strains from the same hospital.

Discussion

Bacterial contamination of hospital facilities was documented intensively around the world, even in hospitals with traditions implemented from reliable infection control and had trustworthy quality management system.

Unfortunately, in Iraq, the routine cleaning and disinfection of hospital wards is often inadequate, partly because of increasing the hospital occupancy density, traffic and human activities during the day, in addition to our hospitals have cleaning policies that vary considerably, even within the same governorate, and rely heavily upon available resources and governmental support. Iraq stills struggle to provide clean water, basic equipment, and trained cleaning staff. Hospital cleaners may receive little or no training for what they are supposed to be doing, and they lack the career advancement experienced by most other professions. Thus, it became clear why the government's public hospitals had such a high level of bacterial contamination in comparison to non-governmental hospitals.We witnessed such dangerous pathogenic prevalence even in CCU and ICU units in Nasseriah hospitals.

In this study, we documented the isolation of 11 different bacterial species. As similar results obtained by many previous studies (17-19), we found that Pseudomonas spp. was the most hospital contaminant bacterial species (30% of all isolated strains) as it consists one of four HAIs microorganisms most frequently isolated in the European Union (20). This bacterium is an opportunistic pathogen that can be found in water distribution systems, water

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containers and as normal flora on the human skin (21). Its ability to survive and spread in hospital environment partly is due to acquisition resistance to commonly used antibiotics and disinfectants(22). These facts makes this species a major life-threatening agent that difficult to avoid in hospitals and was responsible for many outbreaks in different countries (23-25). The second most frequent bacterial type of the city hospital that represents (19%) is Enterobacter spp. According to (CDC), this species, have been considered opportunistic pathogens responsible for 8.6 % of nosocomial infections such as intravascular device-related, and surgical site infections.

In spite the fact of non highly pathogenic *E. coli* strains are common bacteria in human gastrointestinal tract, some of *E. coli* strains such as *E. coli* 0157:H7 are able to produce toxins that induce serious human infections (1). This species comprises 17% of contaminant isolates in Nasseriah.

It is not excluded that the isolated *E. coli* from city's hospitals is a highly pathogenic strain of species, but because of our poor possibility of such a routine survey to classify of isolated pathogenic bacteria into their serotypes.

In fact, *Staph. aureus* which consider as the most reliable indicator of hospital cleaning effectiveness, and known to be a predominant HAIs, because of their ubiquitous human carriage and frequent human traffic (26,27), represented only 12% of all our isolated strains.

Acinetobacter spp., that ranks among the top 10 most common pathogens associated with HAIs, which previously isolated from multiple surfaces and medical equipment (28), formed only 7% of isolated strains in this study. The rest of the contaminant species (*Stenotrophomonas* spp, *Klebsiella* spp, *Burkholderia cepacia*, *Citrobacter* spp and *Micrococcus* spp) were less likely to be present than those previously mentioned, (represented, $\%^{\xi}, \Upsilon$, 1.4%, 0.93%, 0.46%, 0.46% respectively), perhaps because of their susceptibility to disinfectantsthat used or due to short-term staying feature outside human body.

Private hospitals were less contamination than governmental ones, this finding was expected due to selffinancing, limited number of patients and the socioeconomic situation of the patients hospitalized privately. The paying attention to cleanliness and sterilization to achieve good reputation and customer satisfaction, represent different aims in comparison with public hospitals.

Some of the hospitals included in the study are policlinic responsibility. In contrast to specialized hospitals (children and infants hospital). policlinics are often crowded with patients with different diseases, ages and gender, making the contamination more intensive and their unwanted consequences frequent. more Fortunately, four of eleven isolated species in bacterial this study Klebsiella. (Escherichia coli. Acinetobacter, Pseudomonas spp) are classified conditional as (not conventional) pathogens not inevitable by direct contact.

Although, some of isolated bacterial species though be harmless for healthy people, the presence of these pathogens make the hospital microbiologically not a safe environment for risky patients, such as immune-compromised patients or those who are often exposed to

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multiple procedures, invasive devices, those treated in surgical wards and ICUs. In conclusion, we should recall the following recommendations to improve the level of sterilization and cleanliness in the hospitals of thegovernorate:

1. Following rules of universal and standard precautions.

2. Surfaces should be cleanbefore they are disinfected

3. Routineplace cleaning is carried out regularly each morning and in late afternoon, or especiallyduring peak work time.

4. Take precautions to protecthospital environment from airborne andwaterbornetransmission.

5. Use a registered quality of disinfectant that has the best activity against the pathogens, and according with manufacturer'sinstructions.

6. Minimize sharing of medical equipment betweenpatients,and

maximize single patient use devices and equipment.

7. Proper disposal of needles and sharps.

8. Replacement medical devices, tools, and equipment they can be easily contaminated and hardly disinfected by others with opposite features.

9. Preparation sufficient educated and trained staff, ongoing monitoring, and constant upgrading of practice.

10. Establishment of two-way communication between those responsible for cleaning and those responsible for infection control (29).

11. Additional

hygienerequirements,must be applied, such as extensive decontaminationof equipment and surfaces, hand hygiene,use of personal protective equipment,especially in invasive departments...

References

1. S.O. Khelissa, M. Abdallah, C. Jama, C. Faille, N-E Chihib. (2017). Bacterial contamination and biofilm formation on abiotic surfaces and strategies overcome their persistence. JMES, Volume 8, Issue 9, Page 3326-3346

2. KarmenGodičTorkar and SanjaIvić. (2017). Surveillance of bacterial colonisation on contact surfaces in different medical wards.ArhHigRadaToksikol 2017;68:116-126.

3. Lawley TD, Clare S, Deakin LJ, Goulding D, Yen JL, Raisen C, Brandt C, Lovell J, Cooke F, Clark TG, Dougan G. (2010). Use of purified Clostridium difficile spores to facilitate evaluation of health care disinfection regimens. Appl. Environ. Microbiol. 76:6895–6900. <u>http://dx.doi</u> .org/10.1128/AEM.00718-10.

4. Eaton KA, Friedman DI, Francis GJ, Tyler JS, Young VB, Haeger J, Abu-Ali G, Whittam TS. (2008). Pathogenesis of renal disease due to enterohemorrhagic Escherichia coli in germ-free mice. Infect. Immun. 76:3054–3063. http://dx.doi.org/10.1128/IAI.01626-07.

5. Judge C, Galvin S, Burke L, Thomas T, Humphreys H, Fitzgerald-Hughes D. (2013). Search and you will find: detecting extended-spectrum β -lactamase-producing *Klebsiella*pneumoniae from a patient's immediate environment. Infect Control Hosp Epidemiol;34:534-6. doi: 10.1086/670206

Web Site: https://imed.utq.edu.iqEmail:utjmed@utq.edu.iqISSN (Print):1992-92 18, ISSN (Online):1992-92 18

DOI: https://doi.org/10.32792/utq/utjmed/17/1/14/0

6. Kramer A, Schwebke I, Kampf G. (2006). How long do nosocomial pathogens persist on inanimate surfaces? A systematic review. BMC Infect Dis.;6(1):130.

7. Kramer A, Schwebke I, Kampf G.(2006). How long do nosocomial pathogens persist on inanimate surfaces? A systemic review. BMC Infect Dis.;6:130, https://doi.org/10.1186/1471-2334-6-130

8. Vincenzo R., et al (2017).What Healthcare Workers Should Know about Environmental Bacterial Contamination in the Intensive Care Unit.BioMed Research International, Volume 2017, Article ID 6905450, 7 pages https://doi.org/10.1155/2017/6905450

9. Jibrin B. Y., et. al. (2017). Bacterial Contamination of Intensive Care Units at a Tertiary Hospital in Bauchi, Northeastern Nigeria. American Journal of Internal Medicine; 5(3): 46-51

10. Yang D, Zhang Z. (2008). Biofilm-forming *Klebsiella pneumoniae* strains have greater likelihood of producing extended-spectrum _-lactamases. J. Hosp. Infect. 68:369–371. http://dx.doi.org/10.1016/j.jhin.2008.02.001.

11. Dwidjosiswojo Z et al., (2011). Influence of copper ions on the viability and cytotoxicity of *Pseudomonas* aeruginosa under conditions relevant to drinking water environments, Int J HygEnv Health, in press,

12. Tankovic J, et al (1994). Characterization of a hospital outbreak of imipenem resistant *Acinetobacter* baumannii by phenotypic and genotypic typing methods. J. Clin. Microbiol. 32:2677–2681.

13. Neely A, Maley MP, Warden GD. 1999. Computer keyboards as reservoirs for *Acinetobacter*baumannii in a burn hospital. Clin. Infect. Dis. 29:1358–1359. http://dx.doi.org/10.1086/313463.

14. Anna R., Dorota R, Agnieszka C and Małgorzata B.(2017). Bacterial contamination of touch surfaces in Polish hospitals,.MedycynaPracy 2017;68(4):459–467 15. Rusin, P., Maxwell S. &Gerba J.C. (2002). Comparative surface to hand and fingertip to mouth transfer of Gram positive and Gram negative bacteria and phage. Journal of Applied Microbiology. 9,585-592. DOI: 10.1046/j.1365-2672.2002.01734.

16. Oie, S., Hosokawa, I. &Kamiya, A. (2002).Contamination of room handles by methicillin sensitive/methicillin resistance Staphylococcus aureus. Journal Hospital Infection 51,140-143. DOI: 10.1053/jhin.2002.1221. Peutherer JF (Eds.). Medical microbiology. 16th ed. Churchill.

17. Bellido F, Hancock R.(1993). Susceptibility and resistance of P. aeruginosa to antimicrobial agents. In: Campa M, Bendinelli M, Friedman H, eds. Pseudomonaaeruginosa as an opportunistic pathogen. New York, Plenum Press, 1993:321–48.

18. Pena C et al. (2003). An outbreak of carbapenem resistant Pseudomonaaeruginosa in a urology ward. Clinical microbiology and infection, 2003, 9:938–43.

19. OluwatoyosiOladapo, KemebradikumoPondei, Wisdom Olomo, AbisoyeOyeyemi, TamaramiebiNanakede and DimieOgoina.(2017). Bacterial Contamination of the Hospital Environment - The Experience of an Infection Control Team in a Tertiary Hospital in Niger Delta Region of Nigeria. International Journal of tropical disease & health 22(3): 1-9.

Web Site: https://imed.utq.edu.iqEmail:utjmed@utq.edu.iqISSN (Print):1992-92 18, ISSN (Online):1992-92 18

DOI: https://doi.org/10.32792/utq/utjmed/17/1/14/0

20. Zarb P., Coignard B., Griskeviciene J., Muller A., Vankerckhoven V., Weist K., Goossens MM., Vaerenberg S., Hopkins S., Catry B., others., (2012). Euro Surveill17 : 20316.

21. Larson,L.L. and Ramphal , R. (2002). Extended-spectrum betalactamases.SeminRespir Infect., 17(3):189-194.

22. S. Ensayef, S. Al-Shalchi and M. Sabbar. (2009). Microbial contamination in theoperating theatre: a study in ahospital in Baghdad. Eastern Mediterranean Health Journal, Vol. 15, No. 1, p:219

23. Reuter S et al., (2002) "Analysis of transmission pathways of *Pseudomonas*aeruginosa between patients and tap water outlets", Crit Care Med, 10:2222-2228,

24. Blanc DS et al., (2004). "Faucets as a reservoir of endemic *Pseudomonas*aeruginosa colonization/infections in intensive care units", Intensive Care Med, 30: 1964-1968,

25. Vallés J et al., "Patterns of colonization by *Pseudomonas*aeruginosa in intubated patients: a 3-year prospective study of 1.607 isolates using pulsed-field gel electrophoresis with implications for prevention of ventilator-associated pneumonia", Intensive Care Med, 30:1768-1775, 2004.

26. Dancer SJ. (2009). the role of hospital cleaning in the control of hospital acquired infection. J. Hosp. Infect. 73:378–385. <u>http://dx.doi.org/10</u> 1016/j.jhin. 2009.03.030.

27. Dancer SJ. (2008). Importance of the environment in methicillin resistant Staphylococcus aureus acquisition: the case for hospital cleaning. Lancet Infect. Dis. 8:101–113. http://dx.doi.org/10.1016/S1473- 3099(07)70241-4.

28. Hidron AI, Edwards JR, Patel J, et al .(2008) ;for the National Healthcare Safety Network Team and Participating National Healthcare Safety Network Facilities. Antimicrobial-Resistant Pathogens Associated With Healthcare-Associated Infections: Annual Summary of Data Reported to the National Healthcare Safety Network at the Centers for Disease Control and Prevention, 2006–2007. Infect Control HospEpidemiol ;29:996-1011.

29. Stephanie J. Dancer, (2014). Controlling Hospital-Acquired Infection: Focus on the Role of the Environment and New Technologies for Decontamination, Clinical Microbiology Reviews Volume 27 Number 4 p. 665–690.

تحليل نتائح تحري التلوث البكتيري الروتيني في مستشفيات مدينة الناصرية خلال عام ٢٠١٦

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الخلاصة

تلوث المستشفيات هو سبب مهم للعدوى المتأصلة فيها. هدفت هذه الدراسة إلى تقييم حالة التلوث البكتيري في ٢٠ مرفقا مختلفا في اثنتين من المستشفيات الخاصة وخمس مستشفيات عامة في مدينة الناصرية (جنوب العراق) خلال عام ٢٠١٦. ما بين ٥٠ إلى ٢٠٠ مسحة تم جمعها أسبوعيا من كل مرفق، عزلت ٢١٣ سلالة زرع إيجابية، فالفت ١١ نوعا بكتيريا تم الكشف عنها بواسطة الاختبارات البيو كيميائية والإنزيمية. تراوح التلوث بين جنس الزوائف Pseudomonas والإنزيمية. تراوح التلوث بين جنس الزوائف Reedomonas والإنزيمية. تراوح التلوث بين جنس الزوائف Citrobacter spp. وعموما، كانت المستشفيات الخاصة أقل تلوثا من المسلالات المعزولة. وكانت أشهر الصيف والشتاء أقل تلوثا من المستشفيات العامة، والخريفوالمستشفيات متعددة التخصصات اكثر تلوثا من تلك وحيدة والخريفوالمستشفيات متعددة التخصصات اكثر تلوثا من تلك وحيدة التخصص.