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ISSN (Online): 3006-4791

The Antibacterial Activity of Silver Nanoparticles and Detection of the *Fnba* Gene in *S. Aureus* Isolated from Burn Patients

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Abstract: The purpose of the recent study was to profile of biofilm formation gene (fnbA) in *S. aureus* which isolated from burn patients, and to detect the antibacterial activity and MICs of silver nanoparticles against pathogenic *S. aureus* isolated from burn wounds. One hundred and sixty samples from burn patients were collected from August to October 2023 in a burn center in the Thi-Qar province of Iraq. The *S. aureus* was only identified in (35; 21.90%) isolates. Microscopical examination, morphological characterization, diverse biochemical testes, were used to recognize *S. aureus* isolates. The PCR technique results recorded that 42.8% of S. aureus isolates had the *fnbA* gene. The Ag NPs had antibacterial activity against completely *S. aureus* isolates.

Keywords: S. aureus, burn, Ag NPs, MIC, fnbA gene, PCR.

Introduction : Annually, about one million individuals worldwide suffer from burns, making it one of the most prevalent injuries globally (1). Exposure to heat, radiation, electricity, or chemicals can cause damage to the skin, leading to a burn. Severe burns can result in significant complications, including sepsis caused by a bacterial infection, shock due to low blood volume, or the formation of scar tissue due to improper wound healing. Skin injury leads to the death of skin cells, resulting in a significant depletion of body fluids, dehydration, disruption of electrolyte balance, renal failure, and subsequent circulatory collapse. An infection presents a significant risk to the lives of individuals who have sustained burns. As a result of the lack of protection offered by intact layers of the skin, burned skin is very vulnerable to pathogens and other illnesses (1).

The *S. aureus*, was one of the causative agents of infections in burn. It has the ability to gradually and indefinitely establish itself in the nasal cavity of healthy individuals. However, in cases of compromised immune function, the body is susceptible to infecting nearby skin and soft tissues and can even invade deeper tissues and the bloodstream, resulting in systemic infections (2). *Staphylococci* are often detected as pathogens in chronic wounds (3). In these types of wounds, cellular aggregates typically form and are surrounded by a self-produced extracellular matrix (4).

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S. aureus infection involves the presence of several virulence factors that contribute to the processes of colonisation, dissemination, and immune evasion. Staphylococci are responsible for more than 80% of bacterial infections in persons (5). Biofilms are responsible for 60% of burn-related deaths, making them a significant concern in burn cases (6). S. aureus produces two separates yet closely related multifunctional cell wall-anchored (CWA) proteins that specifically attach to the host glycoprotein fibronectin. The fibronectin-binding proteins (*FnBPA* and *FnBPB*) consist of two separate domains (7). One of the first proteins of Gram-positive bacteria to be defined was the fibronectin binding proteins (*FnBPs*) (7). *FnbA* gene facilitate the colonization and infection of the host by S. aureus via adhesion to Fn present in the extracellular matrix of the tissues. Following adhesion, S. aureus is able to invade a variety of non-professional phagocytic mammalian cells, such as epithelial cells, fibroblasts, endothelial cells, osteoblasts (8).

Nanotechnology includes the study and application of science, engineering, and technology at a size of 1 to 100 nanometers. Silver nanoparticles have become a prominent research topic in recent years, among the several types of nanoparticles. (9). Ag NP have great antibacterial characteristics due to their large surface-to-volume ratio, allowing them to effectively combat bacteria even at low concentrations (10). Additionally, they have low costs and have shown minimal harm to cells. Silver nanoparticles have many potentials uses in the field of biomedicine (11). Silver compounds are commonly utilized in medicine to treat burns, wounds, and many infectious diseases (12). Ag+ is combined with microbial DNA and RNA to cause damage, Ag+ inhibits protein translation by destructing 30S ribosomal subunit. And the cell wall synthesis in gram-positive bacteria (13, 14). These nanoparticles exhibit antibacterial properties by adhering to the cell membrane surface, causing disruption of cellular permeability and respiration processes, and inactivating membrane proteins (15). The purpose of this study was to profile of *fnbA* gene in *S. aureus* isolated from burn patients; also, to determine the antibacterial effects and MIC of Ag nanoparticles against *S. aureus*.

Material and methods

Samples Collection and bacterial identification

One hundred and sixty swabs were collected from burn patients, whom were admitted to burn center in the Nasiriyah Hospital during the period from August to October 2023. The specimens were transported in a sterile, leak-proof container to the laboratory, and cultured on blood agar, mannitol salt agar (Biolab, Hungary), Chrom agar (HI media, India), then incubated aerobically for 24 h at 37 °C, the pure isolated colonies of S. aureus were subsequently transferred onto BHI agar medium for the purpose of preservation and conducting additional biochemical tests, which helped confirm the identification of the isolates.

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Antibacterial activity & MIC analysis of Ag nanoparticles against *S. aureus* The antimicrobial activity of Ag NPs was assessed by Agar well diffusion method against *S. aureus* obtained from burn infections. *S. aureus* suspended in D.W, the turbidity generated by the growth was adjusted to match the 0.5 McFarland standard (1.5 X 108 CFU/ml) requires an optical density. A sterile cotton swab was submerged in the suspension and the swab was dipped and used to evenly spread across the whole surface of a Mueller Hinton agar plate. Using a sterile cork-borer, pores with a diameter of 7 mm were formed and filled with 100 µl of Ag NPs with a concentration of $(25\mu g/ml)$ and using the distal water as control. After 24 hours, the Petri plates were placed in an incubator set at 37°C. To determine the antibacterial activity, the growth inhibition zones were measured in millimetres in diameter (16).

To determination of the MICs of the Ag NPs was evaluated using the broth micro-dilution method in a 96-well microtiter plate following the criteria of the Clinical and Laboratory Standards Institute (17). In brief, Bacterial isolates were cultured overnight in BHIB media (Tm, India), then diluted to achieve a turbidity equivalent to 0.5 McFarland Standard using D.W. Fifty μ L of diluted testing substances were added to the wells of a dilution plate, and 50 μ L aliquots of the bacterial suspension were delivered into the wells of this plate. After being incubated at 37 °C overnight, the MIC of the tested agents that stop the observable growth of bacteria were determined and defined as the MIC values.

Extraction of S. aureus DNA and PCR technique

The PrestoTM Mini gDNA Bacteria Kit was utilized to extract the DNA from all isolates of *S. aureus*. The pairs of specific primer of *fnb*A gene as following: forward: 5'- ATC AGC AGA TGT AGC GGA AG -3' and reverse: 5'- TTT AGT ACC GCT CGT TGT CC -3' (18). The total volume of the PCR reaction was 20 μ l, which consisted of the following components: 12.5 μ l of Master Mix (Geneaid/Taiwan), 3 μ liters of bacterial DNA, 1 μ liter of forward and reverse primer for the *fnbA* gene, and the remainder of the volume was filled with nuclease-free water. The program of PCR for *fnbA* gene: initial denaturation at 94°C for 2 min, followed by 30 cycles of denaturation at 94°C for 1 min, annealing at 55°C for 1 min, extension at 72°C for 1 min and final extension for 5min after the last cycle. Agarose gel electrophoresis of PCR product was carried out in 1.4% agarose gel and the presence of a 156bp band indicate a positive result for *fnb* A gene (18).

Ethical approval: The present investigation followed to the ethical standards outlined in the Declaration of Helsinki. The committee of researchers at the Thi-Qar Health Directorate (No. 2023/165 on 8/8/2023) has viewed and approved this study. and verbal consent was obtained from all patients after explaining the purpose of the study.

Statistical analysis: The data of the current study was statistically analysis by using SPSS based in using Chi-square, one-way ANOVA, LSD.

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Results

Bacterial isolation and identification

Out of the 160 from the samples taken from burned individuals in the Thi-Qar province, only 35 (21.90%) of the isolates were able to ferment mannitol and appeared as yellow colonies on the MSA medium, and which grown as green colonies on chrom agar. these colonies were identified as *S. aureus*, as shown in Fig. (1). On the other hand, 53 out of the 160 isolates (33.10%) were identified as other bacterial species. There were 72 samples (45%) that did not show any growth, as depicted in Fig. (2). Completely isolates of *S. aureus* were identified using biochemical assays, including: Catalase, Coagulase and DNase tests which gave positive results for those tests. Statistically, there were a significant difference at (p.<0.001) among bacterial groups.

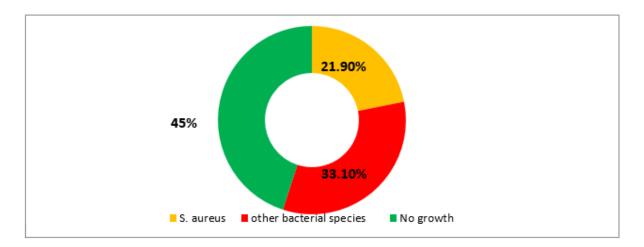


Figure (1): The appearance of *S. aureus* on A-MSA B - chrom agar.

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CalX2= 431, TabX2=7.81, df=3, p. value < 0.001

Figure (2): Distribution of bacterial groups in burn patients.

The current results showed a significant difference at (p. < 0.05; 0.006) of the most isolated *S. aureus* in the patients with third degree of burn (45.71%), followed in the patients with second degree (34.29%), while the lowest isolated *S. aureus* in the patients with first degree of burn (20.0%), as presented in table (1).

Burn Grade	S. Aureus		
	No.	%	
1 st Grade	7	20.0	
2 nd Grade	12	34.29	
3 rd Grade	16	45.71	
Total	35	100	
Cal $X^2 = 10.1$ Tab $X^2 = 5.9$	99 D= 2 P. Value. 0.006		

Antibacterial activity and MIC of Ag NPs against S. aureus isolates

The effect of Ag NPs against completely S. *aureus* isolates recorded that this agent had antibacterial activity, as shown in table (2).

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Table (2) : Antibacterial activity of Ag NPs against all isolates of S. aureus

Inhibition Zone (Mm)	No. Of Isolates	P. Value < 0.001
12-15	3	LSD= 1.76
16	8	
18-20	11	
22-26	9	
27-28	4	

The results of MIC test of Ag NPs against completely *S. aureus* isolates by microdilution method revealed that the turbidity was noticed in microtiter plate wells, 3.125 and 1.562 mg/ml containing Ag nanoparticles indicating the growth of bacteria, and the concentration (6.25 mg/ml) as MIC against *S. aureus* isolates, as revealed in figure (3).

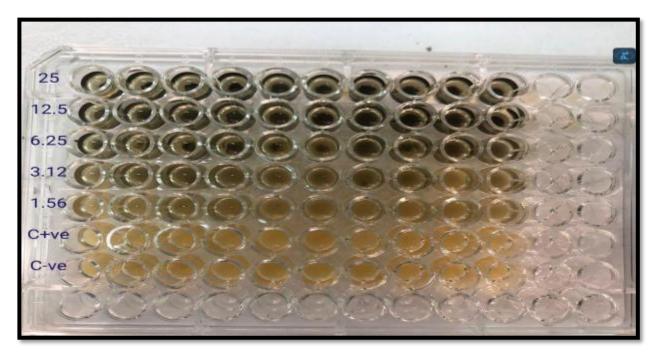


Figure (3): The MIC test of Ag NPs by microdilution method

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Molecular diagnosis

The results of PCR technique showed (15/35; 43%) of *S. aureus* isolates giving positive results for *fnbA* gene. The bands of *fnbA* gene which determined the size of *fnbA* gene, nearly 156bp, as shown in Figure. (4)



Figure (4): Agarose gel electrophoresis of *fnbA* gene amplification, wherever: M : ladder, 3,4,5, 7,8,9,10,11,13 positive results ; 1,2,12,14 negative results.

Discussion

From burn wound swabs from burn patients, approximately 35 (21.90%) exhibited mannitol fermentation on MSA. *S. aureus* is the commonest pathogenic bacteria found in different wound specimens. In recent decades, the prevalence of infections caused by *S. aureus*, namely MRSA strains, in burn centers has emerged as a significant global health concern (19). According to multiple publications, there is an increasing incidence of *S. aureus* infection among burn patients with severe wounds who are admitted to specialized burn centers. Consequently, significant focus has been given to this problem in recent years (20). Continuous surveillance of infections caused

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ISSN (Online): 3006-4791

by *S. aureus* in hospitalized patients is crucial for both preventing and managing these infections, as well as ensuring appropriate and effective treatment (21).

The present results were closely matched with local study conducted in Misan, which found that 18 out of 105 samples (17.14%) were identified as *S. aureus* indicated that *S. aureus* is the primary bacterium found in burn wounds (22).

Studies in the USA found that coagulase-negative *Staphylococci* and *S. aureus* are the most prevalent among their patient groups, with rates of 47.1% and 44.1%, respectively. Gramnegative bacteria (26.5%) have the lowest rate of growth (23).

The present results disagreed with several studies like: the locally study (24) documented that (85/276; 67.460%) identify as methicillin resistance *S. aureus* (MRSA) which collected from burn patients.

The results of PCR technique revealed 43% of the biofilm encoding gene establish in pathogenic *S. aureus* isolated from burn wounds. The examined gene (*fnbA*) gene may be related with increased pathogenicity of pathogenic *S. aureus*. Various microbial factors contribute to the attachment of *S. aureus* to various surfaces. *S. aureus* produces a range of microbial surface components recognizing adhesive matrix molecules that interact with extracellular ligands in the host. (25).

fnbA and *fnbB*, have a crucial impact on the pathogenicity of *S. aureus* due to their capacity to attach to fibronectin and fibrinogen and trigger the uptake of the bacterium by non-professional phagocytes such endothelial cells through integrin-mediated mechanisms (26). Given that biofilm formation enhances resistance to antimicrobial agents and antibiotic resistance, this finding has significant importance. other researcher (27), discovered that the ability of *S. aureus* isolates to respond to antimicrobial mediators generated by mast cells is linked to the increased production of α -hemolysin, fibronectin-binding protein A, and several regulatory mechanisms.

The current results were closely agreed with (28) noticed the results of the genetic analysis of adhesion and biofilm genes as: *fnbA* in (35/83; 42.1%). Also, the research conducted by (29) which found *fnbA* was present in 82.2% of MRSA strains and *fnbB* in 46.7%. Several studies were disagreed with recent results like: (30) revealed that the prevalence of *fnbA* gene was (4.9%); while the study done in Iran observed that the incidence of *fnbA* gene much higher than in the present data (75.7%) (31). Also, other study reported that (81%) of isolates had the *fnbA* gene (32); and (33) recorded that among the examined *S. aureus*, (19/32; 59%) harbored *fnbA* gene.

The results of antimicrobial activity and MIC of Ag nanoparticles against *S. aureus* recorded that the concentration of 25 mg/ml was the best antimicrobial activity due to efficient diffusion in the

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agar medium, causing an inhibition zone, referring to the fact that Ag NPs might cause inhibition of bacterial growth.

The MIC of nanoparticles against examined *S. aureus* was (6.25mg/ml). The scientific community aims to develop a successful alternative to replace available antibiotics that are ineffective due to resistance. Additionally, it serves as an effective alternative to antibiotics due to the increasing challenge of Drug Resistance bacteria. Nanoparticles are being considered as a potential substitute for antibiotics for solving the issue of bacterial resistance (34). The results of study performed by (35,36) showed the MIC concentration against *S. aureus* was (0.039 mg/mL; 0.625 mg/ml), respectively; while, (37) revealed that Ag NPs had high activity with slightly concentration against *S. aureus* isolates as MIC (12.5 μ g/mL).

Conclusion

The silver nanoparticles have potential bacterial effects against pathogenic S. aureus.

Refrences

1-Church D, Elsayed S, Reid O, Winston B, Lindsay R. Burn wound infections. Clinical microbiology reviews. 2006 Apr;19(2):403-34.

2- Kwiecinski JM, Horswill AR. Staphylococcus aureus bloodstream infections: pathogenesis and regulatory mechanisms. Current opinion in microbiology. 2020 Feb 1; 53:51-60.

3- Gjødsbøl K, Christensen JJ, Karlsmark T, Jørgensen B, Klein BM, Krogfelt KA. Multiple bacterial species reside in chronic wounds: a longitudinal study. International wound journal. 2006 Sep;3(3):225-31.

4- Fazli M, Bjarnsholt T, Kirketerp-Møller K, Jørgensen B, Andersen AS, Krogfelt KA, Givskov M, Tolker-Nielsen T. Nonrandom distribution of Pseudomonas aeruginosa and Staphylococcus aureus in chronic wounds. Journal of clinical microbiology. 2009 Dec;47(12):4084-9.

5- Pourhajibagher M, Mahmoudi H, Rezaei-Soufi L, Alikhani MY, Bahador A. Potentiation effects of antimicrobial photodynamic therapy on quorum sensing genes expression: A promising treatment for multi-species bacterial biofilms in burn wound infections. Photodiagnosis and photodynamic therapy. 2020 Jun 1; 30:101717.

6- Goodwine J, Gil J, Doiron A, Valdes J, Solis M, Higa A, Davis S, Sauer K. Pyruvate-depleting conditions induce biofilm dispersion and enhance the efficacy of antibiotics in killing biofilms in vitr o and in vivo. Scientific reports. 2019 Mar 6;9(1):3763.

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ISSN (Online): 3006-4791

7- Speziale P, Pietrocola G. The multivalent role of fibronectin-binding proteins A and B (FnBPA and FnBPB) of Staphylococcus aureus in host infections. Frontiers in microbiology. 2020 Aug 26; 11:2054.

8- Speziale P, Pietrocola G. The multivalent role of fibronectin-binding proteins A and B (FnBPA and FnBPB) of Staphylococcus aureus in host infections. Frontiers in microbiology. 2020 Aug 26; 11:2054.

9- Saravanan M, Barik SK, MubarakAli D, Prakash P, Pugazhendhi A. Synthesis of silver nanoparticles from Bacillus brevis (NCIM 2533) and their antibacterial activity against pathogenic bacteria. Microbial pathogenesis. 2018 Mar 1; 116:221-6.

10- Oves M, Aslam M, Rauf MA, Qayyum S, Qari HA, Khan MS, Alam MZ, Tabrez S, Pugazhendhi A, Ismail IM. Antimicrobial and anticancer activities of silver nanoparticles synthesized from the root hair extract of Phoenix dactylifera. Materials Science and Engineering: C. 2018 Aug 1; 89:429-43.

11-Samuel MS, Jose S, Selvarajan E, Mathimani T, Pugazhendhi A. Biosynthesized silver nanoparticles using Bacillus amyloliquefaciens; Application for cytotoxicity effect on A549 cell line and photocatalytic degradation of p-nitrophenol. Journal of Photochemistry and Photobiology B: Biology. 2020 Jan 1; 202:111642.

12- Avalos A, Haza AI, Mateo D, Morales P. Interactions of manufactured silver nanoparticles of different sizes with normal human dermal fibroblasts. International wound journal. 2016 Feb;13(1):101-9.

13- Blecher K, Nasir A, Friedman A. The growing role of nanotechnology in combating infectious disease. Virulence. 2011 Sep 1;2(5):395-401.

14- Landini P, Antoniani D, Burgess JG, Nijland R. Molecular mechanisms of compounds affecting bacterial biofilm formation and dispersal. Applied microbiology and biotechnology. 2010 Apr;86:813-23.

15- Kwakye-Awuah B, Williams C, Kenward MA, Radecka I. Antimicrobial action and efficiency of silver-loaded zeolite X. Journal of Applied Microbiology. 2008 May 1;104(5):151624.

16- Singh JP, Singh V, Sharma A, Pandey G, Chae KH, Lee S. Approaches to synthesize MgO nanostructures for diverse applications. Heliyon. 2020 Sep 1;6(9).

17- Qu X, Wang S, Qu Y, Wang H, Ye X, Tang L, Xie Q. Antimicrobial Susceptibility Characteristics and Risk Factors Associated with Adult Sepsis in Wenzhou, China. Infection and Drug Resistance. 2022 Jan 1:915-24.

Web Site: <u>https://jmed.utq.edu</u>

Email: <u>utjmed@utq.edu.iq</u>

ISSN (Online): 3006-4791

18- Abraham NM, Jefferson KK. A low molecular weight component of serum inhibits biofilm formation in Staphylococcus aureus. Microbial pathogenesis. 2010 Dec 1;49(6):388-91.

19- Bessa LJ, Fazii P, Di Giulio M, Cellini L. Bacterial isolates from infected wounds and their antibiotic susceptibility pattern: some remarks about wound infection. International wound journal. 2015 Feb;12(1):47-52.

20- Kot B, Sytykiewicz H, Sprawka I. Expression of the biofilm-associated genes in methicillinresistant Staphylococcus aureus in biofilm and planktonic conditions. International journal of molecular sciences. 2018 Nov 6;19(11):3487

21- Kazemzadeh J, Rabiepoor S, Alizadeh S. The quality of life in women with burns in Iran. World journal of plastic surgery. 2019 Jan;8(1):33.

22- Rahim Hateet R. Isolation and Identification of Some Bacteria Contemn in Burn Wounds in Misan, Iraq. Archives of Razi Institute. 2021 Dec 1;76(6):1665-70.

23- Tsurumi A, Que YA, Ryan CM, Tompkins RG, Rahme LG. TNF- α /IL-10 ratio correlates with burn severity and may serve as a risk predictor of increased susceptibility to infections. Frontiers in public health. 2016 Oct 5; 4:216.

24- Degaim ZD, Shani WS, Hamim SS. Virulence factors of Methicillin Resistant Staphylococcus aureus (MRSA) isolated from burn patients. International Journal of Current Microbiology and Applied Sciences. 2015;4(7):898-906.

susceptibility pattern in Tabriz, Iran. Arch Pharm Pract. 2020;11(S1):137-43

25- VázquezSánchez, D., Cabo, M.L., Ibusquiza, P.S., and RodríguezHerrera, J.J., Biofilmforming ability and resistance to industrial disinfectants of Staphylococcus aureus isolated from fishery products, Food Control, 2014, vol. 39, pp. 8–16

26- Violante TL, Haase EM, Vickerman MM. Collagen-binding streptococcal surface proteins influence the susceptibility of biofilm cells to endodontic antimicrobial solutions. Journal of Endodontics. 2013 Mar 1;39(3):370-4.

27- Goldmann O, Tuchscherr L, Rohde M, Medina E. α -hemolysin enhances Staphylococcus aureus internalization and survival within mast cells by modulating the expression of β 1 integrin. Cellular microbiology. 2016 Jun;18(6):807-19.

28- Mohammadi A, Goudarzi M, Dadashi M, Soltani M, Goudarzi H, Hajikhani B. Molecular detection of genes involved in biofilm formation in Staphylococcus aureus strains isolates: evidence from shahid motahari hospital in Tehran. Jundishapur Journal of Microbiology. 2020 Jul 31;13(7).

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ISSN (Online): 3006-4791

29- Mirzaee M, Najar-Peerayeh S, Behmanesh M. Prevalence of fibronectin-binding protein (FnbA and FnbB) genes among clinical isolates of methicillin resistant Staphylococcus aureus. Molecular genetics, microbiology and virology. 2015 Oct; 30:221-4.

30- Chen X, Wu Z, Zhou Y, Zhu J, Li K, Shao H, Wei L. Molecular and virulence characteristics of methicillin-resistant Staphylococcus aureus in burn patients. Frontiers in Laboratory Medicine. 2017 Mar 1;1(1):43-7.

31- Eftekhar F, Rezaee R, Azad M, Azimi H, Goudarzi H, Goudarzi M. Distribution of adhesion and toxin genes in staphylococcus aureus strains recovered from hospitalized patients admitted to the ICU. Archives of Pediatric Infectious Diseases. 2017 Jan 1;5(1).

32- Abad HE, Sadeghi J, Aghazadeh M, Ahangarzadeh Rezaee M, Samadi Kafil H, Ahangar Oskouee M. Frequency of fnbA, fnbB, hla and cna genes in Staphylococcus aureus isolates obtained from blood cultures and their antimicrobial susceptibility pattern in Tabriz, Iran. Arch Pharm Pract. 2020;11(S1):137-43

33- Mater AD, Degaim ZD, Abdulazeez AS. Detection of Fibronectin Binding Protein (fnb A) gene in Staphylococcus aureus isolates. University of Thi-Qar Journal. 2019 Apr 25;14(1):10-6.

34- Morone MV, Dell'Annunziata F, Giugliano R, Chianese A, De Filippis A, Rinaldi L, Gambardella U, Franci G, Galdiero M, Morone A. Pulsed laser ablation of magnetic nanoparticles as a novel antibacterial strategy against gram positive bacteria. Applied Surface Science Advances. 2022 Feb 1;7:100213.

35- Al-Dhabi NA, Mohammed Ghilan AK, Arasu MV. Characterization of silver nanomaterials derived from marine Streptomyces sp. al-dhabi-87 and its in vitro application against multidrug resistant and extended-spectrum beta-lactamase clinical pathogens. Nanomaterials. 2018 Apr 26;8(5):279.

36- Parvekar P, Palaskar J, Metgud S, Maria R, Dutta S. The minimum inhibitory concentration (MIC) and minimum bactericidal concentration (MBC) of silver nanoparticles against Staphylococcus aureus. Biomaterial investigations in dentistry. 2020 Jan 1;7(1):105-9.

37- Viorica RP, Pawel P, Kinga M, Michal Z, Katarzyna R, Boguslaw B. Lactococcus lactis as a safe and inexpensive source of bioactive silver composites. Applied Microbiology and Biotechnology. 2017 Oct;101:7141-53.